pKa Determination of Benzhydrylpiperazine Antihistamines in Aqueous and Aqueous Methanol Solutions

DAVID W. NEWTON **, WALLACE J. MURRAY ‡ , and MICHAEL W. LOVELL $^{\$}$

Received December 14, 1981, from the *Department of Pharmaceutics, College of Pharmacy, [‡]Department of Biomedicinal Chemistry, University of Nebraska Medical Center, Omaha, NE 68105 and the [§]Department of Pharmaceutics, College of Pharmacy, J. Hillis Miller Health Center, University of Florida, Gainesville, FL 32610. Accepted for publication February 3, 1982.

Abstract
The pKa₁ and pKa₂ values of three benzhydrylpiperazine antihistamines, cyclizine (I), chlorcyclizine (II), and hydroxyzine (III), were determined at $24.5 \pm 0.5^{\circ}$ by potentiometric titration in aqueous solution to be 2.16 ± 0.02 and 8.05 ± 0.03 , 2.12 ± 0.04 and 7.65 ± 0.04 , and 1.96 ± 0.05 and 7.40 ± 0.03 , respectively. The pKa₂ values were also determined by titration in seven aqueous methanol solutions in the range of 11.5-52.9% (w/w) methanol. The apparent dissociation constants of I-III in the aqueous methanol solutions, $p_8 Ka_2$, were plotted according to two linear regression equations from which the values in water, $p_{\omega}Ka_2$, were extrapolated. The plotted variables were psKa2 versus methanol concentration (% w/w) and $p_s Ka_2 + \log$ (water concentration, M) versus $1000/\epsilon$, where ϵ is the dielectric constant of the aqueous methanol solution. The maximum difference between pKa_2 and $p_{\omega}Ka_2$ was observed in the case of II where $p_{\omega}Ka_2$ was 5.23% higher. Statistical analysis of the linear regression data obtained from the plots showed that slightly better accuracy (p < 0.13) and correlation (p < 0.16) were obtained, but the precision was essentially equal with both methods. The observed ratio of K_{a1}/K_{a2} in I-III, $2.75 \times 10^5 - 7.76 \times 10^5$, was attributed to solvent- and space-mediated field effects and electrostatic induction between nitrogen atoms in the piperazine ring.

Keyphrases \square Benzhydrylpiperazines—antihistamines, pKa determination in aqueous and aqueous methanol solutions \square Dissociation—pKa determination of benzhydrylpiperazine antihistamines in aqueous and aqueous methanol solutions \square Antihistamines—benzhydrylpiperazines, pKa determination in aqueous and aqueous methanol solutions

Five benzhydrylpiperazine antihistamines are currently marketed in the U.S.: cyclizine (I), chlorcyclizine (II), hydroxyzine (III), meclizine (IV), and buclizine (V).



Although some pKa¹ data for I (1, 2), II (1-3), III (4-7), and IV (6, 8) have been reported (Table I), much of it is based on titrations in 30-50% aqueous alcohols and chlo-

roform-aqueous buffer partition experiments. More recent values of pKa_1 determined potentiometrically for III and IV (6) and spectrophotometrically for III (7) appear to have been conducted more precisely in aqueous solution.

The benzhydrylpiperazine antihistamines are popularly used as prescription and nonprescription dosage forms for their antiemetic, antipruritic, and sedative effects. It was the purpose of this investigation to determine the pKa₁ and pKa₂ values of these drugs for application to various pharmaceutical situations and to evaluate the accuracy of two methods for extrapolating an aqueous pKa estimate from apparent pKa values obtained in aqueous methanol solutions.

BACKGROUND

The low aqueous solubility of nonprotonated amine bases or nondissociated organic acids appears to be the consensus nemesis to determining pKa values of these substances by potentiometric titration. Even the lower limit of $5 \times 10^{-4} M$ for potentiometry (9) is often not achievable. The use of mixed solvent systems, e.g., an alcohol-water and dioxanewater, has been the method most commonly used to surmount this solubility problem. There is little or no disadvantage to this practice if the purpose merely is to compare the relative acidities or basicities of a series of chemical analogs. If, however, the objective is to make judgments pertaining to the aqueous environments of *in vivo* phenomena, the discrepancy between pKa values determined in aqueous and those in aqueous organic solvent solutions could be consequential.

Estimates of aqueous pKa values for inadequately soluble substances have usually been obtained by plots of the apparent pKa values in aqueous organic solvents *versus* solvent concentration (*e.g.*, % w/w) (10, 11):

$$p_s Ka' = [solvent] + p_\omega Ka$$
 (Eq. 1)

where $p_s Ka'$ is the apparent dissociation constant in the aqueous organic solvent system, [solvent] is the organic concentration (% w/w), and $p_\omega Ka$ is the value of $p_s Ka'$ extrapolated to 0% (w/w) organic solvent or 100% (w/w) water. This method was used extensively in pharmaceutical studies during the 1950s and early 1960s (1–3, 12–15). The errors inherent in pKa estimates with this method have been addressed elsewhere (16–18). In 1959, two authors independently proposed a relationship that they rationalized should provide a more accurate estimate of aqueous pKa values

Table I—Acid Dissociation Constant (pKa) Values Reported for Some Benzhydrylpiperazine Antihistamine Drugs

			-
Drug	pKa ₁	pKa ₂	
Cyclizine (I) Chlorcyclizine (II) Hydroxyzine (III) Meclizine (IV)	$\begin{array}{c} 2.54^{a} \\ 2.44^{a}, 2.43^{c} \\ 2.13^{d}, 1.83^{e}, 2.0^{f} \\ 3.1^{h}, 2.05^{i} \end{array}$	$7.92^{a}, 8.16^{b}$ $7.78^{a}, 8.15^{b}, 7.81^{c}$ 7.9^{g} 6.2^{h}	

^a In 50% methanol (Ref. 1). ^b In 30–50% alcohol (Ref. 2). ^c In 20–80% alcohol (Ref. 3). ^d Via partitioning between 0.5 M H₃PO₄ and chloroform (Ref. 4). ^e In aqueous solution at an ionic strength <0.001 (Ref. 6). ^f Spectrophotometrically in aqueous solution (Ref. 7). ^g In unspecified aqueous methanol solutions (Ref. 5). ^h Via partitioning between 0.5 M H₃PO₄ and chloroform (Ref. 8). ⁱ In aqueous solution at an ionic strength of 0.005 (Ref. 6).

¹ The symbol pKa is used throughout in reference to the acid dissociation constant. Formally, K_a is the acid dissociation constant for proton loss by neutral and cationic acids or protonated bases, and pKa = log $(1/K_a)$.

obtained by extrapolation of p_sKa' values in aqueous organic solvent mixtures (19, 20):

$$p_s Ka' + \log [H_2O] = \frac{1}{\epsilon} (e^{2/2.303} akT) - \log B_H$$
 (Eq. 2)

where psKa' is the apparent pKa value in aqueous organic solvent mixtures uncorrected for electrode response, [H₂O] is the water concentration (M), ϵ is the dielectric constant, e is the ionic charge, a is the mean cation-anion ionic diameter, k is the Boltzmann constant, T is the absolute temperature, and B_H is a constant based on the assumption that $[H_3O^+]$ \gg [solvent H⁺]. The values of p_sKa' are corrected by subtracting a constant, δ , which takes into account the variability of glass electrode response in aqueous methanol solutions. This gives the actual pKa value in the mixed solvent system, p_s Ka (21, 22):

$$\mathbf{p}_s \mathbf{K} \mathbf{a} = \mathbf{p}_s \mathbf{K} \mathbf{a}' - \delta \tag{Eq. 3}$$

Thereafter, a plot of $p_sKa + \log [H_2O]$ versus $1/\epsilon$ that is essentially linear may be extrapolated to obtain the ordinate intercept value for p_{ω} Ka, the estimated aqueous pKa when log $[H_2O]$ is 1.74 and $1/\epsilon$ is 0.01273². The linearity of the plots may decrease appreciably when $1/\epsilon > 0.02$ (e.g., \geq 70% w/w methanol). This corresponds to the value of $\epsilon \simeq 50$, below which ionic dissociation begins to diminish with a concomitant increase in ion-pair association and perturbation of the equilibrium between the neutral and ionic species (18, 21).

The original solubility method for determining pKa values (23) has been applied to determining pKa values of acidic, basic, and amphoteric substances with low aqueous solubility (9, 24-26). This method has the advantage of providing thermodynamic pKa estimates when the ionic strength of the samples is low (e.g., <0.0005). Its disadvantages include the failure to achieve equilibrium conditions and separate emulsified or suspended neutral species from the saturated salt solutions in equilibrium with them. Potential errors in solubility estimates resulting from supersaturation of ionic and/or neutral species and the instability of some compounds also jeopardize the accuracy of this method. Another method involving a single pKa titration in an aqueous organic solvent mixture has been proposed, but it was judged by its author to be generally inapplicable to the estimation of p_{ω} Ka values (27).

It is reasonable to expect that the widespread use of organic solventwater mixtures for pKa titrations of substances with intrinsic water solubilities of < 0.0005 M will continue. This is because of historical precedents and the expedient manipulative techniques that can be accomplished using aqueous organic solutions.

EXPERIMENTAL

Materials--Cyclizine hydrochloride (I)³, chlorcyclizine hydrochloride (II)⁴, hydroxyzine dihydrochloride (III)⁵, meclizine dihydrochloride (IV)⁶, and buclizine dihydrochloride (V)⁷ were used as received. Methanol, hydrochloric acid, and potassium hydroxide solutions were ACS analytical reagent grade, and freshly boiled water with a resistivity $\geq 10^7$ ohm cm were used in the solvent systems and titrants.

Procedures-Titrations were conducted in a 25- or 50-ml multinecked glass flask fitted with a thermometer calibrated in 0.1° units. Agitation was provided by a polytef-coated magnetic stirring bar, and pH values were measured with a digital pH meter⁸ equipped with a combination glass electrode⁹. The ambient temperature of the systems varied from 24.0 to 25.0° with an average value of 24.5°. Titrant increments were added from a pipet¹⁰ in 0.25-ml portions.

The $p_s Ka'_2$ values of 0.002 M I-III were determined in seven aqueous methanol solutions in the range of 11.5-52.9% (w/w) with a titrant of 0.020 N KOH prepared in aqueous methanol that contained to within 0.1% (w/w) the same methanol concentration as the solutions of I-III. Values of pKa'2, the apparent constant in aqueous solution, were determined in aqueous 0.001 M I and III titrated with aqueous 0.010 N KOH and in aqueous 0.0005 M II titrated with aqueous 0.005 N KOH. The pKa'₁ values of I and II were obtained from titrations of 0.015 M aqueous solutions with aqueous 0.030 N HCl. The pKa'₁ of III was determined by titrating aqueous 0.015 M III with aqueous 0.030 N KOH.

Activity Corrections-Values of psKa' obtained from titrations of I-III in aqueous methanol solutions were corrected to p. Ka values with Eq. 2 using δ values reported elsewhere (22). The pKa'₂ values of I-III were corrected for activity effects of the K_2 equilibria in Scheme I¹¹ (16, 18, 28–30):

$$HB^{+} + OH^{-} \stackrel{K_{2}}{\longleftrightarrow} B + H_{2}O$$

$$Scheme I$$

$$pKa_{2} = pKa'_{2} - \frac{0.51\sqrt{I}}{1 + 1.6\sqrt{I}}$$
(Eq. 4)

where I is the ionic strength of the solution, and the entire negative term estimates the logarithm of the activity coefficient, $-\log \gamma_i$. A similar activity correction was used for the K_1 equilibria shown in Schemes II and III¹¹ (28):

$$H_{2}B^{+2} + OH^{-} \overleftarrow{\longleftarrow} HB^{+} + H_{2}O$$
Scheme II
$$HB^{+} + H_{3}O^{+} \overleftarrow{\longleftarrow} H_{2}B^{+2} + H_{2}O$$
Scheme III
$$pKa_{1} = pKa'_{1} = \frac{1.5345\sqrt{I}}{1 + 1.6\sqrt{I}}$$
(Eq. 5)

RESULTS AND DISCUSSION

The values of pKa₁ for I-III were determined to be 2.16 ± 0.02 , 2.12 \pm 0.04, and 1.96 \pm 0.05, respectively. The corresponding values of pKa₂ are 8.05 ± 0.03 , 7.65 ± 0.04 , and 7.40 ± 0.03^{12} .

The plots of the data for $p_s Ka_2$ versus methanol (% w/w) are shown in Fig. 1 and that for $p_s Ka_2 + \log [H_2O]$ versus 1000/ ϵ in Fig. 2 for I-III. Least-squares regression lines were plotted for all I-III data sets to permit comparison of the accuracy and precision of paKa2 extrapolations derived from each set of ordinate and abscissa coordinates. The ϵ values in Fig. 2 were calculated for each aqueous methanol solution from a linear regression equation derived from data for 10-60% (w/w) methanol (31):

$$\epsilon = -0.4523C + 78.9307 \tag{Eq. 6}$$

where C is the methanol concentration $(\% \text{ w/w})^{13}$. The linear regression data for the plots in Figs. 1 and 2 are summarized in Tables II and III, respectively.

A comparison of R values for I-III (Tables II and III) shows that, based on the Student's t test (32), there was a better linear correlation of the p_s Ka₂ titration data plotted according to Eq. 2 (p < 0.16). There was, however, essentially no difference (p < 0.001) in the precision of the plots of Eqs. 1 and 2 based on a comparison of $S_{y,x}$ values (Tables II and III). The accuracy of $p_{\omega}Ka_2$ values for I-III, *i.e.*, the differences from pKa_2 (Tables II and III), was also better by Eq. 2 than Eq. 1 plots (p < 0.13). The slightly improved linear correlation of the Eq. 2 plot may explain, in part, why it has been advocated instead of Eq. 1 (16, 18). Furthermore, the unreported linear regression data for plots of $p_s Ka_2 + \log [H_2O]$ versus methanol (% w/w) were virtually identical with those reported in Table III. This would be expected from the nearly linear relationship expressed by Eq. 6 (31).

Doubts about the accuracy of p_{ω} Ka values extrapolated from aqueous organic solvent mixtures containing >20% solvent have been expressed elsewhere (16-20); curves shaped like hockey sticks have resulted from Eq. 1 plots. One contributing cause of this could be plotting erroneous high values of p_s Ka' uncorrected for δ (Eq. 3). The value of δ ranged from 0.02-0.12 in these titrations, increasing with methanol concentration. Failure to have corrected p_sKa' to p_sKa values would have caused progressive decreases in the slopes of Eq. 1 plots. The resulting horizontal inclinations, i.e., the hockey stick appearance, at high methanol concentrations would have produced greater inaccuracy in the extrapolated

² The values of log [H₂O] and $1/\epsilon$ correspond to [H₂O] = 55.3 *M* and ϵ = 78.54, ³ Intervalues on log [12:0] and 1/s correspond to [12:0] = 55 spectively, for water at 25°.
 ³ Lot 9C0080, Burroughs Wellcome Co., Greenville, N.C.
 ⁴ Lot 8J0053, Burroughs Wellcome Co., Greenville, N.C.
 ⁵ Lot 1753-27EA, 99.6%, Pfizer Inc., Brooklyn, N.Y.
 ⁶ Lot 2F268-81EA, 100.0%, Pfizer Inc., Brooklyn, N.Y.
 ⁷ Lot AN52650, Stuart Pharmaceuticals, Wilmington, Del.
 ⁸ Model 105

 ⁸ Model 125, Corning Scientific Products, Medfield, Mass.
 ⁹ No. 476050, Corning Scientific Products, Medfield, Mass.
 ¹⁰ Eppendorf Model 4700, Brinkmann Instruments Inc., Westbury, N.Y.

 $^{^{11}}$ B, HB⁺, and H_2B⁺² refer to the nonprotonated, monoprotonated, and dipro-

¹² Values of pKa₁, pKa₂, and p₈Ka₂ represent the means \pm standard deviations of seven values in each set of ten titrations. ¹³ The correlation coefficient, *R*, is 0.9997.



Figure 1—Plot of p_sKa versus methanol (% w/w) for cyclizine (I), chlorcyclizine (II), and hydroxyzine (III). Key: (\bullet) I; (\Box) II; (\circ) III.

 $p_{\omega}Ka_2$ values. Therefore, the I–III p_sKa_2 data were calculated for 37.7% (w/w) \leq methanol \leq 52.9% (w/w) and 16.2 \leq 1000/ $\epsilon \leq$ 18.2 (Table IV). Statistical analysis showed no difference in the accuracy and precision of I–III $p_{\omega}Ka_2$ values (p < 0.001) and the linear correlation of Eq. 1 and Eq. 2 plots (p < 0.001) between the data in Tables II and IV or Tables III and IV. These findings pertain only to I–III under the conditions stated, and they do not warrant general advocation of extrapolating $p_{\omega}Ka$ values from p_sKa data obtained in >35% (w/w) methanol.

A literature search yielded one similar study of some phenothiazine derivatives (33). Those plots were based on fewer data (mostly three points) obtained in a higher methanol concentration range (mostly $\geq 40\%$) and lacked the statistical data necessary for comparing pKa with p_{ω} Ka values derived from Eqs. 1 and 2. The plots apparently consisted of empirically connecting the sets of coordinates rather than calculating least-squares regression lines (33).

The magnitude of error in calculating the percent of monoprotonated I–III that would result from using $p_{\omega}Ka_2$ values to estimate otherwise



Figure 2—Plot of $p_{s}Ka + log [H_2O]$ versus $1000/\epsilon$ for cyclizine (I), chlorcyclizine (II), and hydroxyzine (III). Key: (\bullet) I; (\Box) II; (O) III.

unreported pKa_2 values for I–III was determined for pH 7.4. The error ranged from 0.72% for I (Table III) to 17.70% for II (Table II) (34):

Percent HB⁺ = $100/[1 + 10^{(pH-pKa_2)}]$ (Eq. 7)

Percent
$$HB_{\omega}^{+} = 100/[1 + 10^{(pH-p_{\omega}Ka_2)}]$$
 (Eq. 8)

Error,
$$\% = |Eq. 7 - Eq. 8|$$
 (Eq. 9)

where HB⁺ is the monoprotonated conjugate acid of I–III. From Eqs. 7 and 8, as the absolute values of the quantities $pH - pKa_2$ and $pH - p_{\omega}Ka_2$ increase, the value of Eq. 9 decreases.

A specific reason for the differences between $p_{\omega}Ka_2$ and pKa_2 values of I-III obtained by either Eq. 1 or Eq. 2 is not readily apparent. The fact that the titrants and I-III solutions were prepared in the same aqueous methanol concentrations, *i.e.*, within 0.1% (w/w), precluded stoichiometric errors in precision resulting from the nonadditivity of volumes and negative heats of solution. The variability in solvation phenomena

Table II—Linear Regression Data for Plots of Equation 1	Tal	ble	II-	Linear	Regression	Data for	Plots of	Equation 1	8
---------------------------------------------------------	-----	-----	-----	--------	------------	----------	----------	------------	---

Drug	Equation, $p_s Ka_2^b =$	p _∞ Ka₂ ^c	pKa2 ^d	Absolute Value Difference, % ^e	R ^f	$S_{y,x}{}^{g}$
I	$\begin{array}{r} -0.011C + 8.067 \\ -0.012C + 8.052 \\ -0.006C + 7.332 \end{array}$	8.07	8.03	0.50	0.983	0.032
II		8.05	7.65	5.23	0.995	0.015
III		7.33	7.40	0.95	0.972	0.023

^a When 11.5% (w/w) \leq methanol \leq 52.9% (w/w). ^b This refers to straight line plots for I-III (Fig. 1) where C is the methanol concentration (% w/w), ^c The value of p_sKa₂ extrapolated to 0% (w/w) methanol. ^d The value of pKa'₂ determined in aqueous solution and corrected for $-\log \gamma_i$ (Eq. 4). ^e Calculated from $|(pKa_2 - p_{\omega}Ka_2)|$ (100/pKa₂). ^f Correlation coefficient. ^g Standard error of the estimate of p_sKa_2 based on the methanol concentration.

Table III—Linear Regression Data for Plots of Equ	uation 24	a
---------------------------------------------------	-----------	---

Drug	Equation, $p_s Ka_2 + \log [H_2O]^b =$	p _∞ Ka₂ ^c	pKa2 ^d	Absolute Value Difference, % ^e	R!	$S_{y,x}^{g}$
I II III	$\begin{array}{c} -0.164 \; (1000/\epsilon) + 11.838 \\ -0.164 \; (1000/\epsilon) + 11.785 \\ -0.121 \; (1000/\epsilon) + 10.584 \end{array}$	8.01 7.96 7.30	8.03 7.65 7.40	$0.25 \\ 4.05 \\ 1.35$	0.997 0.997 0.994	0.022 0.019 0.023

^a When $14.5 \le 1000/\epsilon \le 18.2$. ^b This refers to straight line plots for I-III (Fig. 2) where $[H_2O]$ and ϵ are the molarity of water and dielectric constant, respectively, of the aqueous methanol solvents. ^c This is the extrapolated value of $p_s Ka_2$ obtained when $\log [H_2O] = 1.743$, where 55.3 is the molarity of water at 25°, is subtracted from the quantity ($p_s Ka_2 + \log [H_2O]$) when the latter is calculated for $1000/\epsilon = 12.73$ where $\epsilon = 78.54$ for water at 25°. ^d Footnote d, Table II. ^e Footnote e, Table II. ^f Footnote f, Table II. ^e Standard error of the estimate of $p_s Ka_2 + \log [H_2O]$ based on $1000/\epsilon$ values.

Т	able	: IV	-Linear	Regression	Data for	Plots of	Equations	1 and 2 ^a

$\mathbf{p}_{\omega}\mathbf{K}\mathbf{a}_{2}$			Difference, % ^d		\mathbf{R}^{e}		$S_{\mathbf{v},\mathbf{x}}^{f}$		
Drug	Eq. 1 ^b	Eq. 2 ^c	pKa ₂	Eq. 1	Eq. 2	Ēq. 1	Eq. 2	Eq. 1	Eq. 2
I	8.25	8.09	8.03	2.74	0.75	0.971	0.990	0.029	0.027
III	7.40	7.32	7.65 7.40	4.18 0	2.88	0.985	0.997	0.014 0.022	0.012 0.029

^a Where 37.7% (w/w) \leq methanol \leq 52.9% (w/w) and 16.2 \leq 1000/ $\epsilon \leq$ 18.2, respectively, for Eqs. 1 and 2. ^b Footnote c, Table II. ^c Footnote c, Table III. ^d Footnote d, Table II. ^c Footnote g, Table III. ^d Footnote g, Table II.

and electrolytic equilibria of I–III in aqueous methanol solutions may partially account for the discrepancies between the pKa₂ and $p_{\omega}Ka_2$ values (16–19, 21, 22, 33). For example, different solvation mechanisms are possible with III than with I or II because the hydroxyethoxyethanol substituent, R₁ on III, is both a hydrogen bond acceptor and donor. Furthermore, the R₂ chloro substituent on II would increase its partition coefficient over that of I (35, 36). In fact, 0.002 *M* II was inadequately soluble in 11.5% (w/w) methanol while titrating its $p_s Ka_2$, which is attributed to the enhanced hydrophobicity conferred by the chloro group, R₂ (35, 36). The $p_s Ka_1$ values of 0.0005 *M* IV and V were obtainable only in solvents containing $\geq 47.8\%$ (w/w) methanol. As noted earlier, titration data derived from the latter solutions would be poorly applicable to $p_{\omega} Ka_1$ estimates (18, 21).

Finally, it remains to be determined why the ratios of K_{a1}/K_{a2} for I–III show a variation of 2.75×10^5 to 7.76×10^5 . The comparable ratio of K_{a1}/K_{a2} for piperazine (VI) was reported to be 1.38×10^4 (37)¹⁴. There are four main types of electrochemical effects that can account for these K_{a1}/K_{a2} ratios: (a) mesomerism or resonance in aromatic compounds, (b) inductive effects of substituents in aliphatic and aromatic compounds, (c) field effects such as intramolecular hydrogen bonding, and (d) molecular symmetry or statistical effects (38–42).

Because the piperazine ring is nonaromatic, mesomerism does not explain the observed K_{a1}/K_{a2} ratios. Secondly, the inductive effects of the R_1 and benzhydryl substituents on the piperazine group in I-III account for a maximum difference between K_{a1} and K_{a2} of ~62 as seen from the quotient of $(K_{a1}/K_{a2})_{1/}(K_{a1}/K_{a2})_{VI}$. It has been shown that electron-withdrawing (-I) groups have a marginal influence on K_a values of acids when the -I and ionogenic groups are separated by more than two -CH₂ residues (38, 40). These inductive effects account primarily for the differences between either K_{a1} or K_{a2} values of I-III. Thirdly, the statistical symmetry factor in VI and its analogs accounts for only a fourfold difference in K_{a1} and K_{a2} values (38-40, 42).

Solvent- and space-mediated field effects and inductive forces between nitrogen atoms of the piperazine ring appear to be the most plausible explanation for the large K_{a1}/K_{a2} ratios in I–III, VI, and similar compounds. Electrostatic attraction between the protonated and neutral nitrogen atoms of the piperazine ring has been alluded to (43), and it has been studied by conformational analysis using molecular orbital methods. (44). However, field effects mediated through solvent molecules may also contribute to these large observed ratios.

CONCLUSIONS

Aqueous pKa₁ and pKa₂ values for I and II, and pKa₂ for III, apparently heretofore unpublished, were determined. These pKa₂ values of I-III were also obtained in aqueous solutions of 11.5–52.9% (w/w) methanol. There was no substantial difference in the precision of $p_{\omega}K_2$ extrapolated from linear regression plots of p_sK_2 data according to Eqs. 1 and 2. However, Eq. 2 did yield slightly better correlation of the plotted coordinates. The overall accuracy of $p_{\omega}Ka_2$ compared to pKa₂ values for I-III also was better according to Eq. 2 plots (p < 0.13). The maximum difference observed between any pKa₂ and $p_{\omega}Ka_2$ value occurred in the case II using Eq. 1 by which $p_{\omega}Ka_2$ was 5.23% higher.

The large ratios of K_{a1}/K_{a2} for I-III and piperazine are attributed to field effects and intramolecular electrostatic attraction or induction in the piperazine ring.

REFERENCES

(1) R. Baltzly, S. DuBreuil, W. S. Ide, and E. Lorz, J. Org. Chem., 14, 775 (1949).

(2) P. B. Marshall, Br. J. Pharmacol., 10, 270 (1955).

(3) N. G. Lordi and J. E. Christian, J. Am. Pharm. Assoc., Sci. Ed., 45, 300 (1956).

(4) B.-A. Persson, Acta Pharm. Suec., 5, 335 (1968).

(5) A. Pardo, S. Vivas, F. Espana, and J. I. Fernandez-Alonso, Afinidad, 29, 640 (1972).

(6) S. Lukkari, Farm. Aikak., 80, 161 (1971); through Chem. Abstr., 75, 41145t (1971).

(7) J. Tsau and N. DeAngelis, in "Analytical Profiles of Drug Substances," vol. 7, K. Florey, Ed., Academic, New York, N.Y., 1978, p. 325.

(8) B.-A. Persson and G. Schill, Acta Pharm. Suec., 3, 281 (1966).

¹⁴ The pKa₁ and pKa₂ values of VI are 5.68 and 9.82, respectively (37).

(9) R. H. Levy and M. Rowland, J. Pharm. Sci., 60, 1155 (1971).

(10) M. Mizutani, Z. Physik. Chem., 116, 350 (1925); through: Chem. Abstr., 19, 3052 (1925).

(11) N. F. Hall and M. R. Sprinkle, J. Am. Chem. Soc., 54, 3469 (1932).

(12) E. B. Leffler, H. M. Spencer, and A. Burger, *ibid.*, **73**, 2611 (1951).

(13) P. J. A. W. Demoen, J. Pharm. Sci., 50, 350 (1961).

(14) L. G. Chatten and L. E. Harris, Anal. Chem., 34, 1495 (1962).

(15) E. R. Garrett, J. Pharm. Sci., 52, 797 (1963).

(16) L. Z. Benet and J. E. Goyan, ibid., 56, 665 (1967).

(17) A. Albert and E. P. Serjeant, in "The Determination of Ionization Constants," 2nd ed., Chapman and Hall, London, England, 1971, pp. 39-40.

(18) R. F. Cookson, Chem. Rev., 74, 5 (1974).

(19) T. Shedlovsky, in "Electrolytes," B. Pesce, Ed., Pergammon, New York, N.Y., 1962, pp. 146-151.

(20) M. Yasuda, Bull. Chem. Soc. Japan, 32, 429 (1959); through Chem. Abstr., 54, 2894e (1960).

(21) R. G. Bates, M. Paabo, and R. A. Robinson, J. Phys. Chem., 67, 1833 (1963).

(22) K. C. Ong, R. A. Robinson, and R. G. Bates, Anal. Chem., 36, 1971 (1964).

(23) H. A. Krebs and J. C. Speakman, J. Chem. Soc., 1945, 593.

(24) I. Setnikar, J. Pharm. Sci., 55, 1190 (1966).

(25) C. C. Peck and L. Z. Benet, ibid., 67, 12 (1978).

(26) U. G. Hennig, R. E. Moskalyk, L. G. Chatten, and S. F. Chan, *ibid.*, **70**, 317 (1981).

(27) J. Peeters, ibid., 67, 127 (1978).

(28) A. Albert and E. P. Serjeant, in "The Determination of Ionization Constants," 2nd ed., Chapman and Hall, London, England, 1971, pp. 28-32.

(29) P. J. Niebergall, in "Remington's Pharmaceutical Sciences," 16th ed., A. Osol, Ed., Mack Publishing, Easton, Pa., 1980, pp. 228–233.

(30) J. Kielland, J. Am. Chem. Soc., 59, 1675 (1937).

(31) P. S. Albright and L. J. Gosting, *ibid.*, 68, 1061 (1946).

(32) L. K. Randolph and J. L. Ciminera, in "Remington's Pharmaceutical Sciences," 16th ed., A. Osol, Ed., Mack Publishing, Easton, Pa., 1980, pp. 111-114.

(33) A. Hulshoff and J. H. Perrin, Pharm. Acta Helv., 51, 65 (1976).

(34) A. Albert and E. P. Serjeant, in "The Determination of Ionization Constants," 2nd ed., Chapman and Hall, London, England, 1971, p. 104.

(35) A. Leo, C. Hansch, and D. Elkins, Chem. Rev., 71, 525 (1971).

(36) S. H. Yalkowsky, in "Design of Biopharmaceutical Properties through Prodrugs and Analogs," E. B. Roche, Ed., American Pharmaceutical Association, Washington, D.C., 1977, pp. 392, 397, 407.

(37) G. Schwarzenbach, B. Maissen, and H. Ackerman, Helv. Chim. Acta, **35**, 2333 (1952); through Chem. Abstr., **47**, 4238c (1953).

(38) J. March, in "Advanced Organic Chemistry: Reactions, Mechanisms and Structure," 2nd ed., McGraw-Hill, New York, N.Y., 1977, pp. 238–243.

(39) J. Hine, in "Structural Effects on Equilibria in Organic Chemistry," Wiley, New York, N.Y., 1975, pp. 1-3, 29-53.

(40) R. P. Bell, in "The Proton in Chemistry," 2nd ed., Cornell University, Ithica, N.Y., 1973, pp. 86–102.

(41) E. S. Gould, in "Mechanism and Structure in Organic Chemistry," Holt, Rinehart and Winston, New York, N.Y., 1959, pp. 199–212.

(42) S. W. Benson, J. Am. Chem. Soc., 80, 5151 (1958).

(43) L. L. Ciaccio, S. R. Missan, W. H. McMullen, and T. C. Grenfell, Anal. Chem., 29, 1670 (1957).

(44) W. J. Murray and D. W. Newton, in "Abstracts," vol. 11, no. 1, APhA Academy of Pharmaceutical Sciences, St. Louis, Mo., Mar.-Apr., 1981, p. 100.

ACKNOWLEDGMENTS

Presented to the Medicinal Chemistry and Pharmacognosy Section at the 31st National Meeting of the APhA Academy of Pharmaceutical Sciences, Orlando, Fla., November 16, 1981.

The authors thank the Burroughs Wellcome Co., Pfizer Inc., and ICI Americas Inc. for generous donations of cyclizine and chlorcyclizine hydrochlorides, hydroxyzine and meclizine dihydrochlorides, and buclizine dihydrochloride, respectively. They are grateful to Mrs. Donna Earnshaw for preparing the manuscript and to Donald L. Goode, Pharm.D., for performing many titrations.